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Research Article

Epiphyseal Cartilage and Subchondral Bone of Rat Tibia as Early Targets of Diabetes Type I

Pulitano Manisagian, Gisela Estefanía, Lewicki, Marianela, Villarino, Mariano Enrique and Ubios, Ángela Matilde

Department of Histology and Embryology, School of Dentistry, University of Buenos Aires, Ciudad Autónoma de Buenos Aires, Argentina

Correspondence should be addressed to: Pulitano Manisagian, Gisela Estefanía; giselapulitano@gmail.com

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Abstract

Little is known about the early response of bone metabolism to hyperglycemia, or transient hyperglycemia, caused by poor compliance with treatment or inadequate control of diabetes in patients with type 1 diabetes. There are no studies comparing early diabetes-related alterations in endochondral ossification with those occurring through time. In order to histomorphometrically evaluate the growth cartilage and subchondral bone of male Wistar rat tibia shortly after experimentally inducing type 1 diabetes, thirty-five male Wistar rats were divided into 5 groups: Group A: control; Group B: induction of diabetes; euthanasia at one week; Group C: induction of diabetes plus insulin treatment; euthanasia at one week; Group D: induction of diabetes; euthanasia at six weeks; Group E: induction of diabetes plus insulin treatment; euthanasia at six weeks. Diabetes was induced by single intraperitoneal injection of 60mg/kg body weight of streptozotocin dissolved in citrate buffer pH 4.0. Two days after inducing diabetes, treatment with subcutaneously injected human insulin was initiated, according to protocol. The following parameters were evaluated: Tibial subchondral bone volume; trabecular area; bone surface; trabecular number; trabecular thickness. Tibial subchondral bone volume and epiphyseal cartilage width were significantly lower in both experimental groups as compared to controls. Insulin treatment resulted in values similar to controls. As shown by the microarchitecture of subchondral trabecular bone, all parameters were lower in diabetic animals, except for trabecular width which was similar to that observed in controls. The alteration in bone metabolism manifests in long bones very early in the course of diabetes, suggesting it would be advisable to analyze bone response as soon as possible following diagnosis of the disease, in order to prescribe treatments aimed at preventing further alterations in bone architecture.

Keywords: Early diabetes-related changes; late diabetes-related changes; endochondral ossification, bone microarchitecture.

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Introduction

Alterations of bone metabolism are a common long term complication in diabetic patients, including those treated with insulin. Although there is a controversy in the literature regarding the implication of such alterations, it is generally accepted that type 1 diabetes is associated with osteopenia, as stated by Tuominen et al. (1999), and type 2 diabetes is associated with increased bone mineral density, as shown by Barrett-Connor and Kritz-Silverstein (1996) and Stolk et al. (1996). Nevertheless, Auwerx et al. (1998), Muñoz-Torres et al. (1996), and Tuominen et al. (1999), found bone mineral density to decrease, and Wakasugi et al. (1993) reported it remained unchanged.

The impact of type 1 diabetes on bone metabolism involves several mechanisms. As described by Baylink et al. (1993) and Chiarelli et al. (2004), a decrease in local levels of IGF-1 would cause a decrease in bone formation. According to Lu et al. (2003), another factor that compromises osteoblast differentiation is the decreased expression of transcription factors involved in its regulation, such as Cbfa1/Runx-2 and Dlx5. In addition, it is well documented that insulin is essential to osteoblast function, as well as to chondrogenesis and collagen synthesis, as reported in the Editorial of Journal Bone; Diabetes and fragility fractures - A burgeoning epidemic? Bone, (2008).

Bouillon (1991) found streptozotocininduced type 1 diabetic rats to show a decrease in bone remodeling. Goodman and Hori (1984) and Inzerillo and Epstein (2004) also observed a decrease in the number of osteoclasts.

It is well known that insulin treatment reverses, at least in part, the effects of hyperglycemia, so that strict insulin therapy normalizes glycemia levels. A study by Hofbauer *et al.* (2007) demonstrated the anabolic effect of insulin on bone tissue. Nevertheless, there are bone alterations that manifest even in diabetic patients on strict insulin regimens, and even at an early age, as reported by Little is known about the early response of bone metabolism to hyperglycemia caused by poor compliance with treatment or inadequate control of diabetes. Moreover, there are no studies comparing the early and long term effects of diabetes on endochondral ossification. Our hypothesis was that diabetes-induced bone alterations manifest soon after the onset of the disease, even when they have not been noticed by the patient or have not become clinically evident.

Based on the above, the aim of the present study was to histomorphometrically evaluate the growth cartilage and subchondral bone of male Wistar rat tibia shortly after experimentally inducing type 1 diabetes.

Materials and Methods

Thirty-five male Wistar rats were assigned to one of five groups of 7 rats each.

Group A: Control animals

The animals in the experimental groups were treated according to the following protocols:

Group B: Induction of diabetes; euthanasia at one week.

Group C: Induction of diabetes plus insulin treatment; euthanasia at one week.

Group D: Induction of diabetes; euthanasia at six weeks.

Group E: Induction of diabetes plus insulin treatment; euthanasia at six weeks.

All the animals received rat chow and water "ad libitum".

At the onset of the experiment, the animals in groups A, D and E were 6 weeks old, and those in groups B and C were 11 weeks old. The average weight at baseline was 115 g in groups A, D, and E and 240 g in groups B and C. All the animals were euthanized at

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the age of 12 weeks. The average final body weight was 268 g in Group A, 215 g in Group B, 255 g in Group C, 180 g in Group D, and 315 g in Group E.

Diabetes was induced by administration of a single intraperitoneal injection of 60 mg/kg of body weight of streptozotocin (STZ) (SIGMA-ALDRICH, Inc., Saint Louis, USA) dissolved in freshly prepared citrate buffer, pH 4.0. The animals in the control group received an equivalent volume of citrate buffer. Twenty-four hours postinjection, blood samples were obtained from all the animals in order to determine blood glucose levels, according to the glucose-oxidase method, using Accu-Chek Sensor Comfort test strips (Roche Ltda. products, Santiago, Chile) in an Accu-Chek Sensor (Roche Diagnostics GMBH, Mannheim, Germany).

All the animals showing blood glucose levels above 250mg/dl were included in the study as diabetic rats. Likewise, control animals were checked to confirm their blood glucose levels were below 120 mg/dl. Glycemia was monitored weekly in non-treated experimental diabetes rats, and every 48 h in the insulin-treated experimental diabetes groups. The body weight of each animal was recorded every 24 h using an Ohaus CS2000 digital weighing scale (Ohaus Corp., N.J., USA).

Two days after inducing diabetes, the animals in experimental groups C and E were initiated on treatment with subcutaneously injected human insulin NPH: recombinant DNA origin (Humulin N, Eli Lilly and Compañía de México, S.A de C.V. Mexico D.F.) 100 IU of Protamina Zinc/ml. The daily insulin dose ranged from 6-20 IU/kg body weight. Subcutaneous administration was performed at the level of the neck fold between 7:00 am and 10:00 am. Treatment requirements were adjusted daily according periodic to quantitative determinations of blood glucose levels and daily follow up of body weight.

All the animals were the same age when euthanized. Euthanasia was performed by

intraperitoneal injection of a solution of 150 mg/kg body weight of sodium thiopental (Pentovent, 49 Richmond Laboratories, Buenos Aires, Argentina) and 3 mg/kg body weight of acepromazine maleate (Acedan, Holliday-Scott S.A., Buenos Aires, Argentina).

Immediately following euthanasia, the tibiae were resected, dissected, fixed in formalin buffer (ph 7.0), decalcified in 10% EDTA (ph 7.0) for 30 days, and embedded in paraffin. Longitudinal sections of the proximal end of the paraffin embedded tibiae were obtained and stained with hematoxylin-eosin.

Histomorphometry

The histomorphometric study was performed based on the stereologic principles stated by Weibel (1967), which allow evaluating the three-dimensional microstructure of a histological tissue section based on the analysis of the twodimensional information observed in the slice. The method by Dalle Carbonare published in the review on Bone Microarchitecture (2005) was used here.

The histological sections were digitally photographed using a Carl Zeiss Axioskop 2 photomicroscope light (Carl Zeiss Mikroskopic, Jena, Germany) equipped with a 5X objective and a Canon Powershot A640 digital camera with a resolution of 10.0 Megapixels and optical zoom of 4X (Canon Inc., China). The histomorphometric study was performed using Pro-Plus 4.0 image analysis software.

Epiphyseal cartilage determinations: The parameters were measured in the 1600 μ m area corresponding to the mid-epiphyseal region. Epiphyseal cartilage width was measured at 9 sites 200 μ m apart and perpendicularly to the chondro epiphyseal junction (Figure 1). Proliferative cartilage width, hypertrophic cartilage width, and total cartilage width were determined. Values corresponding to each animal were averaged and expressed in μ m.

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Figure 1: The Lines Show the Sites Where Epiphyseal Cartilage Width was Determined. As Shown in the Figure, Measurements were Performed from the Middle Caudal Region of the Epiphyseal Cartilage

Subchondral bone determinations: Measurements were performed in the subchondral bone tissue in a $5.2 \times 10^6 \mu m^2$ area (Figure 2); this was the area considered to define tissue volume (TV), and used to measure all histomorphometric bone parameters.



Figure 2: Areas Where the Histomorphometric Parameters were Measured. Note the Areas are Located in the Middle Region below the Epiphyseal Cartilage

The following parameters were studied:

- Tibial subchondral bone volume (Bone Volume/Tissue Volume-BV/TV: %), defined as the fraction of total volume corresponding to trabecular bone.
- Trabecular area (TA: mm²) Defined as the sum of the areas containing bone trabeculae.
- Bone surface: Defined as the sum of the perimeter of all trabeculae (Bone Surface-BS: mm)
- Trabecular Number (Tb.N: 1/mm)
- Trabecular Thickness (Tb.Th; μm)
- Trabecular Separation (Tb.Sp:µm)

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The results were statistically analyzed using one way analysis of variance and Bonferroni's multiple comparisons test. Statistical significance was set at a value of p < 0.05.

Results

Histomorphometric Determinations of the Epiphyseal Cartilage

Total epiphyseal cartilage width was significantly lower in experimental animals

compared to controls. The difference was more marked in late-stage than in earlystage diabetic rats, and was less marked in insulin-treated than in non-insulin treated animals. Proliferative cartilage and hypertrophic cartilage widths were also lower in experimental animals compared to matched controls, but only the differences in proliferative cartilage reached statistical significance (Figure 3).



Figure 3: Total Growth Cartilage Width was Significantly Lower in All the Experimental Groups as Compared to the Control Group; Proliferative Cartilage and Hypertrophic Cartilage Widths were Also Lower in Experimental Animals Compared to Controls, but Only the Differences in Proliferative Cartilage Width Reached Statistical Significance

Histomorphometric Parameters of Bone:

 Tibial subchondral bone volume (BV/TV): Bone volume was lower in all the experimental groups compared to controls; the difference only reached statistical significance in Group D. Bone volume was higher in insulin-treated diabetic rats than in non-treated rats, though the difference only reached statistical significance when comparing Groups D and E (Figure 4).

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Figure 4: Tibial Subchondral Bone Volume was Significantly Lower in Diabetic Animals in Group D than in Controls. Bone Volume was Significantly Lower in Group D than in Insulin-Treated Group E, and Did Not Differ Significantly between the Latter and Controls (Group A)

- Trabecular area: The results followed the same pattern as the bone volume results, and are shown in Figure 5.



Figure 5: Trabecular Area was Significantly Smaller in Group D as Compared to Group A (Controls) and Group E, and was Not Significantly Different between the Two Latter Groups

- Number of trabeculae: The results followed a similar pattern to that of trabecular area, as shown in Figure 6.

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- Trabecular microarchitecture: Although trabecular separation was found to increase in all experimental groups as compared to controls, only the increase in group D was statistically significant compared to the increases observed in the remaining groups. Trabecular Thickness in all the experimental groups was similar to that observed in controls. Both parameters are shown in Figure 7.





Histological images are shown in Figure 8.

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Figure 8: Frontal Sections of the Proximal End of the Tibia. (A): Control; (B): Induction of Diabetes; Euthanasia at One Week; (C): Induction of Diabetes Plus Insulin Treatment; Euthanasia at One Week; (D): Induction of Diabetes; Euthanasia at Six Weeks; (E):
Induction of Diabetes Plus Insulin Treatment; Euthanasia at Six Weeks. Growth Cartilage Width was Lower in Diabetic Animals in Groups (B) and (D) than in Controls (A), and was Similar in Groups (C) and (E) and Controls (A). There were Fewer Subchondral Bone Trabeculae in Diabetic Groups (B) and (C); Trabeculae were More Spaced, and Shorter than Control Trabeculae (A). Trabeculae in Insulin-Treated Diabetic Group (E) were Similar to those of Controls (A)

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Discussion

The results of the present study showed that tibial subchondral bone volume and epiphyseal cartilage width were significantly lower in short term and long term diabetic rats than in control rats. In both cases insulin treatment prevented these alterations, and values were similar to controls.

The experimental model used here to study early diabetes-related events has been validated by several authors, including Olubadewo and Spitzer (2003) who indeed used it to study immunologic alterations, and Wald et al. (1998), who used it to study cholinergic neurotransmission. It was used in the present work to study early diabetesrelated changes in a long bone, before systemic compensatory mechanisms that tend to normalize the metabolic alteration are triggered. According to Villarino et al. (2010), the model in alveolar bone is also useful to analyze responses to occasional transient hyperglycemia that can occur in diabetic patients when control of diabetes inadequate or compliance is with treatment is poor. This is the first study to the demonstrate occurrence of microstructural alterations in the epiphyseal cartilage and bone in rats shortly after the diabetic induction, and that rapid administration of insulin prevents the decrease in bone volume, suggesting its preventive effect against the aforementioned alterations.

The late changes observed in bone in nontreated animals in the present study are similar to those reported by Hamada et al. (2007). The latter authors observed a decrease in static and dynamic parameters of bone activity and in bone volume, which reversed with insulin therapy. Our results are also in keeping with those of Thrailkil et al. (2005), who reported a significant decrease in bone formation in chronic diabetic mice, which also reversed with insulin therapy. Clinical and experimental studies conducted by Bouillon (1991) and Thrailkill et al. (2005) demonstrated decreased osteoblast function in diabetes. Duarte et al. (2005) studied biochemical, histomorphometrical and densitometric alterations in diabetic rats up to 120 days after the induction of diabetes, and concluded that osteopenia ensues is related to the duration of the illness.

Vestergaard (2007), As stated by Janghorbani et al. (2007) and Miao et al. (2005), it is known that type 1 diabetic patients are at a 6 or 7-fold higher risk of fracture than the non-diabetic hip population. Our experimental studies on trabecular microarchitecture at the level of subchondral bone, involving evaluation of trabecular number, width, and separation, showed that all the parameters studied here were lower in diabetic animals, except for trabecular thickness, which remained unchanged. The alterations in bone microarchitecture explain the increased risk of long bone fracture in diabetic patients.

Although Strotmeyer et al. (2006) and Mastrandrea et al. (2008) observed decreased BMD (Bone Mineral Density) in patients with type 1 diabetes and suggested that the decrease may partly explain the increased fracture risk, according to Vestergaard (2007) it would not be the only causal factor involved in diabetes-related skeletal fragility. As explained by Armas et al. (2011), measurements of bone quality, such as variations in bone remodeling rates and defects in bone microarchitecture like those shown in the present study, may provide a better explanation for the observed increase in fracture risk.

The structural alterations observed in long bones in untreated long term diabetic rats are the consequence of early changes in the growth cartilage that manifest as a decrease in cartilage width, which in turn leads alterations to in hone microarchitecture. This finding is due to a decrease in trabecular number and an increase in trabecular separation, despite trabecular thickness remaining unchanged. It is therefore essential to highlight the importance of early detection and prompt treatment of the disease, particularly in the pediatric population.

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Conclusions

The alterations in bone metabolism manifest in long bones very early in the course of diabetes. It would seem advisable to analyze bone response as soon as possible once the disease has been diagnosed, in order to initiate treatments aimed at preventing major alterations in bone architecture.

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The study was approved by the Ethics Committee of the School of Dentistry, University of Buenos Aires (N^o FOUBA. UBACYT 2011-2014-3), and was performed in keeping with the National Institutes of Health Guidelines for the Care and Use of Laboratory Animals (NIH publication 85-123 Rev. 2010).

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