

Journal of Respiratory Medicine Research and Treatment

Vol. 2014 (2014), Article ID 162245, 50 minipages. DOI:10.5171/2014.162245 www.ibimapublishing.com

Copyright © 2014. Attapon Cheepsattayakorn and Ruangrong Cheepsattayakorn. Distributed under Creative Commons CC-BY 3.0

Research Article

Dengue-Virus Infection: Lung Involvement, Clinical Implications, and Associated Human Leukocyte Antigens

Authors

Attapon Cheepsattayakorn¹ and Ruangrong Cheepsattayakorn²

¹10th Zonal Tuberculosis and Chest Disease Centre, Chiang Mai, Thailand, 10th Office of Disease Prevention and Control, Department of Disease Control, Ministry of Public Health, Thailand

²Department of Pathology, Faculty of Medicine, Chiang Mai University, Chiang Mai, Thailand Received date: 6 February 2014

Accepted date: 15 May 2014

Published date: 4 August 2014

Academic Editor: Hirofumi Akari

Cite this Article as: Attapon Cheepsattayakorn and Ruangrong Cheepsattayakorn (2014), " Dengue-Virus Infection: Lung Involvement, Clinical Implications, and Associated Human Leukocyte Antigens ", Journal of Respiratory Medicine Research and Treatment, Vol. 2014 (2014), Article ID 162245, DOI: 10.5171/2014. 162245

Abstract

Four known dengue virus serotypes have been identified, DENV-1, DENV-2, DENV-3, and DENV-4. About 50 millioninfected persons occurs each year. Adult primary infections with DENV-1 and DENV-3 usually result in dengue fever while some outbreak with DENV-2 infections have been predominantly subclinical. Dengue pulmonary complications include pulmonary infiltration, pleural effusion, noncardiogenic pulmonary edema and respiratory failure while massive hemoptysis can occur. Several human leukocyte antigens class I and II alleles are associated with the development of dengue disease.

Keywords: Dengue, Lung, HLA

Introduction

This disease is caused by dengue virus (DENV) that belongs to the family Flaviviridae, genus Flavivirus, and is transmitted to humans by Aedes mosquitoes, mainly Aedes aegypti [Martina et al., 2009]. Four serotypes of viruses have been identified; DENV-1, DENV-2, DENV-3, and DENV-4 [Martina et al., 2009]. An estimated 50 million-infected people occur each year and more than 2.5 billion people are being at risk of infection [Guha-Sapir et al., 2005], but the simultaneous worldwide distribution of the risk of dengue virus infection and its public health burden are poorly understood [Bhatt et al., 2013]. Epidemic with high incidences of dengue hemorrhagic fever have been linked to primary infection with DENV-1 followed by infection with DENV-2 or DENV-3,

whereas it indicated that the longer the interval between primary and secondary infections, the higher the risk of developing severe disease [Guzma'n et al., 2003-Ong et al., 2007]. The relationship between DENV-2 and dengue severity is controversial [Di'az-Quijano et al., 2012]. However, adult dengue infections are frequently accompanied by a tendency for severe hemorrhage [UNICEF, UNDP, World Bank, accessed January 8, 2014] and can be life-threatening when infections occur in patients withchronic diseases such as asthma and diabetes [Kouri et al., 1987-Lee et al., 2006]. The aim of this study is to review the clinical implications and associated human leukocyte antigens including lung pathological mechanisms and involvement in dengue disease.

Associated Human Leukocyte Antigens, Lung Involvement and Clinical Implications

Lungs of dengue cases, particularly with severe disease, present with mononuclear inflammatory infiltrates, hyperplasia of alveolar macrophages, and hyaline membrane formation with concomitant hypertrophy of type II pneumocytes [Po'voa et al., 2014]. Dengue virus antigens with virus replication are also identified in type II pneumocytes and pulmonary vascular endothelium [Po'voa et al., 2014]. These pulmonary pathological features can contribute to pulmonary edema, pulmonary hemorrhage, adult respiratory distress syndrome, and pulmonary tissue damages [Po'voa et al., 2014]. Several human leukocyte antigens (HLA) class I alleles, female sex, AB blood group, a single-nucleotide

polymorphism in the tumor necrosis factor gene, and a promoter variant of the DC-SIGN receptor gene are the host factors that increase the risk of severe dengue disease [Stephens et al., 2002-Kalayanarooj et al., 2007]. Notably, the first outbreak in the Americas occurred in 1981, which coincided with the introduction of the possibly more virulent DENV-2 Southeast Asian genotype whereas the less virulent indigenous DENV-2 genotype was already circulating in the region [Kouri et al., 1987, Rico-Hesse et al., 1990-Rodriguez-Roche et al., 2005]. Age has been demonstrated to influence the disease outcome following a secondary infection with heterologous DENV [Guzma'n et al., 2002]. In Asia, the risk of severe disease is greater in children than in adults, in contrast to the Americas [Cologna et al., 2003, Leitmeyer et al., 1999]. Nevertheless, the finding that dengue hemorrhagic

fever or dengue shock syndrome is noted primarily in a relative small percentage of secondary DENV infections and to a much lesser extent in primary infections although with virulent strains it indicates that host factors must be critical determinants of severe DENV disease development [Martina et al., 2009]. There is evidence that DENV antigen is present in the pulmonary vascular endothelium [Jessie et al., 2004], whereas liver is the organ commonly involved in human DENV infections including mouse model [Paes et al., 2005, Seneviratne et al., 2006].

Glucose-6-phosphate dehydrogenase deficiency which is highly prevalence among the African population [Nkhoma *et al.*, 2009] can cause abnormal cellular redox, therefore affecting the production of hydrogen peroxide, superoxide, and

nitric oxide indicating oxidative stress [Wu et al., 2008]. Viral proliferation and virulence by increasing viral receptors on target cells or increasing viral particles is known to be affected by oxidative stress [Wu et al., 2008], therefore, glucose-6-phosphate dehydrogenase deficiency may contribute to the increased replication of DENV in monocytes [Nkhoma et al., 2009]. Several HLA class I alleles (-A*01, -A*0207, -A*24, -B*07, -B*46, -B*51) [Stephens et al., 2002, Loke et al., 2002, Zivna et al., 2002] and HLA class II alleles (-DQ*1, -DR*1, -DR*4) [LaFleur et al., 2002, Polizel et al., 2004] are associated with the development of dengue hemorrhagic fever. Additionally, a recent study demonstrated that there was significantly higher frequency of HLA-A*33 allele in dengue fever patients, HLA-A*0211 allele in dengue hemorrhagic fever cases compared to healthy controls and dengue fever cases.

respectively [Alagarasu et al., 2013]. The frequency of HLA-*B*18* and *HLA-Cw*07* alleles were significantly higher in DENV-infected cases compared to controls [Alagarasu et al., 2013]. The combined frequency of HLA-Cw*07 with HLA-DRB1*07/*15 genotype was significantly higher in dengue hemorrhagic fever cases as compared to dengue fever cases and controls, but the frequency of combination of HLA-Cw*07 allele without HLA-DRB1*07 allele was significantly higher in dengue fever cases compared to controls [Alagarasu et al., 2013]. This study result indicates that HLA-A*33 allele may be associated with the development of dengue fever, whereas HLA-B*18 and HLA-Cw*07 alleles may be associated with symptomatic dengue infection requiring hospitalization [Alagarasu et al., 2013]. A previous study demonstrated that HLA-A*0207 and HLA-B*51 alleles were associated with

dengue hemorrhagic fever in patients having secondary DENV-1 or DENV-2 infection only and children with *HLA-A*24* allele were more likely to develop dengue hemorrhagic fever [Malavige *et al.*, 2004]. After secondary dengue infections, *HLA-B*44*, *-B*62*, *-B*76*, and *-B*77* alleles revealed that they protect against development of clinical disease [Malavige *et al.*, 2004].

Rathakrishnan and colleagues conducted a study in 504 Chinese and Indian Malaysian populations, who aged 14 and above, which demonstrated that *HLA-A*24*, *HLA-A*33*, and *HLA-B*57* alleles were positively associated with patients with warning signs of dengue disease or severe dengue disease [Rathkrishnan *et al.*, 2014]. *HLA-A*03* allele may be protective in both Chinese and Indian Malays, whereas *HLA-A*33* allele may be a predictive marker for the development of severe dengue disease [Rathkrishnan et al., 2014]. Cardozo et al demonstrated the results of their study of susceptibility of dengue virus serotype 3 among 95 patients that HLA-DOA1*05:01 and HLA-DRB1*11 alleles could be possible resistance factors to dengue virus serotype 3 infection, whereas HLA-DQB1*06:11 and HLA-DRB1*15 alleles may act as susceptible factors [Cardozo et al., 2014]. Alencar and colleagues conducted a study of a cohort of dengue patients in Brazil and demonstrated that HLA-B*44, HLA-B*50, HLA-DR*16 alleles were associated with increased susceptibility to dengue hemorrhagic fever, particularly serotype 3, whereas HLA-B*07 and HLA-DR*13 alleles were associated with resistance to secondary dengue infection with DENV-3 [Alencar et al., 2013]. Monteiro et al conducted a

retrospectively case (dengue hemorrhagic fever)-control (dengue fever) study among Brazilians during 2002-2008 and revealed that *HLA-A**01 allele was associated with increased susceptibility to dengue hemorrhagic fever, whereas *HLA-A*31* allele was associated with resistance to the development of dengue hemorrhagic fever [Monteiro et al., 2012]. Brown and colleagues conducted a study among Jamaicans and demonstrated that HLA-A*24 and HLA-DRbeta5*01/02 alleles were associated with increased susceptibility to dengue infection, whereas HLA-A*23, HLA-*CW**04, *HLA-DObeta**02, *HLA-DObeta**03, and *HLA-DObeta**06 alleles were associated with protection to dengue infection [Brown et al., 2011]. A previous study in Sri Lanka demonstrated that *HLA-A*31* allele was associated with dengue shock syndrome during secondary dengue infection,

while *HLA-A*24* and *HLA-DRB1*12* alleles were associated with the development of dengue hemorrhagic fever during primary dengue infection [Malavige *et al.*, 2011].

Appanna et al's study demonstrated that HLA-B*18 and $HLA - B^{*53}$ alleles were increased in patients with dengue hemorrhagic fever, whereas HLA-A*03 allele was decreased [Appanna et al., 2010]. Falco'n-Lezama and colleagues conducted a study among Mexican patients with dengue fever (23) and dengue hemorrhagic fever (16) in comparison to 34 healthy controls, and revealed that HLA-DQB1*0202 and HLA-DQB1*0302 alleles were associated with the development of dengue fever and dengue hemorrhagic fever, respectively [Falco'n-Lezama et al., 2009]. A study in 228 ethnic Thais with dengue fever and 142 patients with dengue

hemorrhagic fever, which was further classified by disease severity according to the World Health Organization (WHO) criteria, aged 3-14 years, demonstrated that HLA-B*48, HLA-B*57 and HLA-DPB1*0501 alleles were associated with the development of secondary dengue hemorrhagic fever [Vejbaesya et al., 2009]. Lan and colleagues conducted a study in Vietnam by gathering 211 patients with dengue hemorrhagic fever and 418 patients with dengue shock syndrome during 2002-2005, and revealed that HLA-A*24 allele was associated with the development of dengue hemorrhagic fever and dengue shock syndrome, whereas HLA-DRB1*0901 allele had protective effect against the dengue shock syndrome caused by DENV-2 [Lan et al., 2008]. Sierra et al demonstrated that HLA-DRB1 polymorphism was

associated with protective effect against the development of dengue hemorrhagic fever [Sierra *et al.*, 2007].

Clinical findings in early febrile stage include fever, headache, malaise, rash, body pain, and later develop pleural effusion [UNICEF, UNDP, World Bank, accessed January 8, 2014, Likitnukul et al., 2004], both lower lobes infiltration [Likitnukul et al., 2004], bilateral perihilar edema [Ali et al., 2010], ascites, bleeding, thrombocytopenia (platelet < 100,000 per mm³), hematocrit > 20%, and clinical warning signs such as restlessness, severe and continuous abdominal pain, persistent vomiting and a sudden reduction in body temperature associated with profuse perspiration, advnamia (vigor or loss of strength) and sometimes fainting which can be indicative of shock due to plasma extravasation [UNICEF,

UNDP, World Bank, accessed January 8, 2014]. Wang and colleagues conducted a study in 661 Taiwanese patients diagnosed with dengue fever according to the clinical presentations and laboratory examination results and revealed that pleural effusion was the most common chest roentgenographic presentations (31.4% of the total chest roentgenograms and 57.4% of the abnormal chest roentgenograms), followed by pulmonary infiltration only (23.3% of the total chest roentgenograms and 42.6% of the abnormal chestroentgenograms), while small pleural effusion (less than 2 intercostal spaces) was the predominate type among the chest roentgenograms presented with pleural effusion and in all abnormal chest roentgenograms [Wang et al., 2007].

Additionally, pulmonary infiltration only and small pleural effusion were the major presentations in dengue hemorrhagic fever patients with abnormal chest roentgenographic features [Wang et al., 2007]. A previous study conducted in 100 patients in Yemen with seropositivity of dengue confirmed by reverse-transcriptase polymerase-chain-reaction method, demonstrate the chest presentations as the following: 1) adult respiratory distress syndrome in patients with dengue fever, dengue hemorrhagic fever, and dengue shock syndrome was 0%, 19.4%, and 53.3%, respectively; 2) pulmonary hemorrhage in patients with dengue fever, dengue hemorrhagic fever, and dengue shock syndrome was 0%, 21.4%, and 6.6%, respectively; 3) unilateral pneumonitis in patients with dengue fever, dengue hemorrhagic fever, and dengue shock syndrome was 0%, 0%, and 0%, respectively; 4) bilateral pneumonitis in

patients with dengue fever, dengue hemorrhagic fever, and dengue shock syndrome was 0%, 9.5%, and 6.6%, respectively; 5) pleural effusion in patients with dengue fever, dengue hemorrhagic fever, and dengue shock syndrome was 0%, 7.14%, and 0%, respectively; 6) more than one presentation in patients with dengue fever, dengue hemorrhagic fever, and dengue shock syndrome was 0%, 38.1%, and 33.3%, respectively [Mohamed *et al.*, 2013].

Asghar *et al* conducted a study of pulmonary manifestations in 76 confirmed dengue-hemorrhagic-fever patients, age ranking from 14 to 80 years, and demonstrated that right pleural effusion, bilateral pleural effusion, left pleural effusion, bilateral pneumonia, right sided pneumonia, and left sided pneumonia was 17.1%, 13.2%, 1.3%, 3.9%, 1.3%, and 0%, respectively [Asghar et al., 2011]. A study of chest computerized tomography (CT) in 29 Brazilian patients who fulfilled the World Health Organization case definition by diagnoses of dengue fever (9 patients) and severe dengue disease (20 patients) showed that abnormal CT findings were identified in 58.62% (12 with severe disease, 5 with dengue fever), pleural effusion in 55.17% (11 with severe disease, 5 with dengue fever, 13 with bilateral effusion, 3 with rightsided effusion), large pleural effusion in 4 patients with severe disease [Rodrigues et al., 2014]. Large pleural effusion was not identified in patients with dengue fever [Rodrigues et al., 2014]. Ground-glass opacity was the most common finding of lung parenchymal involvement that was noted in 8 patients (5 with severe disease, 3 with dengue fever) and followed by lung consolidation (6 patients (4 with severe

disease, 2 with dengue fever)) [Rodrigues et al., 2014]. Interlobar septal thickening and pulmonary nodules with no specific distribution were detected in 2 patients (1 with severe disease, 1 with dengue fever, and 2 with severe disease, respectively) [Rodrigues et al., 2014]. One case with dengue fever and another case with severe disease demonstrated mild interlobar septal thickening that located predominantly in the upper lobes and lower lung zone, respectively [Rodrigues et al., 2014]. Only one case with intermediately severe disease demonstrated peribronchovascular interstitial thickening in the middle and lower zones [Rodrigues et al., 2014]. No patient with dengue fever showed pulmonary nodules [Rodrigues et al., 2014]. There was no specific axial distribution [Rodrigues et al., 2014]. The chest extent of disease tended to be greater

in patients with severe disease than in those with dengue fever, but this difference was not statistically significant [Rodrigues et al., 2014]. Transudative pleural effusions that were mostly detected in dengue patients are largely due to imbalances in oncotic and hydrostatic pressures in the thoracic cavity because these effusions are ultrafiltrates of plasma [Wang et al., 2007]. Generally, the identification of a transudative pleural effusion indicates that the pleural membranes are not diseased and that pleural fluid accumulation is caused by systemic (non-pleural, non-lung) factors affecting the formation and absorption of pleural fluid [Wang et al., 2007]. Dengue disease must be excluded from two syndromes related to hantavirus, hemorrhagic fever with renal syndrome (HFRS) in Eurasia and hantavirus pulmonary syndrome (HPS) in the Americas [Duchin et al., 1994-Vapalahti *et al.*, 2003]. HPS is typically characterized by acute noncardiogenic pulmonary edema and circulatory shock, whereas fever, hemorrhagia, and acute renal failure are hallmark findings in HFRS [Rasmuson *et al.*, 2011].

Laboratory diagnosis of DENV infection includes virus isolation, serodiagnostic tests (MAC-ELISA, IgG ELISA, IgG : IgM ratio, neutralization assay), nucleic acid amplification tests (real-time PCR, reverse-transcriptase PCR, nucleic acidsequence based amplification assay (NASBA)), and antigen detection (NS1 antigen and antibody detection) [UNICEF, UNDP, World Bank, accessed January 8, 2014]. DENV complications include massive hemorrhage or hemoptysis, disseminated intravascular coagulation, non-cardiogenic pulmonary edema, respiratory failure, and finally develop

multiple organ failure [UNICEF, UNDP, World Bank, accessed January 8, 2014]. The chest roentgenographic presentations are significantly correlated with the laboratory findings, particularly white blood cell counts, platelet counts, activated partial thromboplastin time, and serum alanine aminotransferase and albumin levels [Wang *et al.*, 2007].

In uncomplicated dengue cases, treatment is only supportive, but in cases with prolonged or recurrent dengue shock, intravenous fluids should be administered carefully according to dosage and age to prevent pulmonary edema [UNICEF, UNDP, World Bank, accessed January 8, 2014]. DENV control and prevention strategies include vector control and vaccine development [UNICEF, UNDP, World Bank, accessed January 8, 2014]. Current approaches to vaccine development involve

using deoxyribonucleic acid vaccine, chimeric viruses using vellow fever vaccine, subunit vaccine, inactivated viruses, attenuated viruses, and attenuated dengue viruses as backbones[Guirakhoo et al., 2006-Edelman et al., 2003]. An Acambis/Sanofi Pasteur vellow fever-dengue chimeric vaccine is in advanced Phase II testing in children in Thailand [UNICEF, UNDP, World Bank, accessed January 8, 2014]. A possible licensed vaccine will be available in less than 10 years [UNICEF, UNDP, World Bank, accessed January 8, 2014].

Conclusions

Presently, dengue is a global health threat and is tropically endemic or epidemic. Better use of currently available measures and interventions should be made while we wait for novel diagnostics, novel vaccines, and novel antivirals. Recently, several partnerships such as the Asia-Pacific Dengue Prevention Partnership and the Innovative Vector Control Consortium have come into existence and receive funding from the Bill and Melinda Gates Foundation, regional Development Banks and the private sector. These partnerships are collaborating with the WHO and national governmental organizations to develop novel tools and strategies to improve diagnostics, clinical therapies, and successful novel vaccines. The most common chest presentation is pleural effusion. To date, the number of known HLA alleles with susceptible effects on the development of dengue infection or severe disease are more than the number of known HLA alleles with protective effects, thus, the development of novel protective measures

against dengue virus infection are urgently needed worldwide, particularly in the tropical regions. The summary of associated HLA alleles and chest roentgenographic presentations in dengue disease is demonstrated in table 1.

Please see Table 1 in the PDF version.

References

1.Martina, B.E.E., Koraka, P. and Osterhaus, A.D.M. (2009), " Dengue virus pathogenesis : an integrated view," *Clinical Microbiology Reviews*, 22 (4), 564-581.

2.Guha-Sapir, D. and Schimme, r B (2005), "Dengue fever : new paradigms for a changing

epidemiology," Emerging Themes in Epidemioogyl, 2 (1), 1.

3.Guzma'n, M.G. and Kouri, G (2003), "Dengue and dengue hemorrhagic fever in Americas: lessons and challenges," *Journal of Clinical Virology*, 27 (1), 1-13.

4. Bhatt, S., Gething, P.W., Brady, O.J., Messina, J.P., Farlow, A.W., Moyes, C.L., *et al* (2013), "The global distribution and burden of dengue," *Nature*, 496 (7446): 504-507.

5. Halstead, S.B (2007), "Dengue," *Lancet* 370 (9599), 1644-1652.

6. Ong, A., Sandar, M., Chen, M.I. and Sin, L.Y (2007), "Fatal dengue hemorrhagic fever in adults during a dengue epidemic in Singapore," *International Journal of Infectious Diseases*, 11 (3),263-267.

7. Di'az-Quijano F.A. and Waldman E.A. (2012), "Factor associated with dengue mortality in LatinAmerica and Caribbean, 1995-2009 : an ecological study," *American Journal of Tropical Medicine and Hygiene*, 86 (2), 328-334.

8. UNICEF, UNDP, World Bank, WHO. Evaluating diagnostics-Dengue : a continuing global threat. (Retrieved January 8, 2014), http://www.nature.com/reviews/micro 9. Kouri, G.P., Guzma'n, M.G. and Bravo, J.R (1987), "Why dengue hemorrhagic fever in Cuba? 2.An integral analysis, *Transactions of the Royal Society of Tropical Medicine and Hygiene*, 81 (5), 821-823.

10. Halstead, S.B., Nimanitaya, S. and Cohen, S.N (1970), " Observations related to pathogenesis of dengue hemorrhagic fever : Relation of disease severity to antibody response and virus recovered," *Yale Journal of Biology and Medicine*, 42 (5), 311-328.

11. Lee, M.S., Hwang, K.P., Chen, T.C., Lu, P.L. and Chen, T.P (2006), "Clinical characteristics of dengue and dengue hemorrhagic fever in a medical center of southern Taiwan during the 2002 epidemic," *Journal of*

Microbiology, Immunology and Infection, 39 (2), 121-129.

12. Po'voa, T.F., Alves, Ada.M.B., Oliveira, C.A.B., Nuovo, G.J., Chagas, V.L.A. and Paes, M.V.P (2014), "The pathology of severe dengue in multiple organs of human fatal cases : histopathology, ultrastructure and replication," *PLoS ONE*, 9 (4), e83386. DOI :10.1371/jpournal.pone.0083386

13. Stephens, H.A., Klaythong, R., Sirikong, M., Vaughn, D.W., Green, S., Kalayanarooj, S., *et al* (2002), "HLA-A and HLA-B allele associations with secondary dengue virus infections correlate with disease severity and the infecting viral serotype in ethnic Thais," *Tissue Antigens*, 60 (4), 309-318.

14. LaFleur, C., Granados, J., Vargas-Alarcon, G., Ruiz-Morales, J., Villarreal-Garza, C., Hiqueral, L., *et al* (2002), "HLA-DR antigen frequencies in Mexican patients with dengue virus infection : HLA-DR4 as a possible genetic resistance factor for dengue hemorrhagic fever," *Human Immunology*, 63 (11), 1039-1044.

15. Loke, H., Bethell, D.B., Phuong, C.X., Dung, M., Schneider, J., White, N.J., *et al* (2002), "Strong HLA class I-restricted T-cell responses in dengue hemorrhagic fever : a double-edged sword ? "*Journal of Infectious Diseases*, 184 (11), 1369-1373.

16. Sakuntabhai, A., Turbpaiboon, C., Casade'mont, I., Chuansumrit, A., Lowhoo, T., Kajaste-Rudnitski, A., *et al* (2005), " A variant in CD209 promoter is associated with severity of dengue disease, *"Nature Genetics*, 37 (5), 507-513. 17. Fernandez-Mastre, M.T., Gendzekhadze, K., Rivas-Vetencourt, P. and Layrisse, Z (2004), "TNF- α -308A allele, a possible severity risk factor of hemorrhagic manifestation in dengue fever patients," *Tissue Antigens*, 64 (4), 469-472.

18. Kalayanarooj, S., Gibbons, R.V., Vaughn, D., Green, S., Nisalak, A., Jarman, R.G., *et al* (2007), "Blood group AB is associated with increased risk for severe dengue disease in secondary infections," *Journal of Infectious Diseases*, 195 (7), 1014-1017.

19. Rico-Hesse, R (1990), "Molecular evolution and distribution of dengue viruses type 1 and 2 in nature," *Virology*, 174 (2), 479-493.

20. Rico-Hesse, R., Harrison, L.M., Salas, R.A., Tovar, D., Nisalak, A., Ramos, C., *et al* (1997), "Origins of dengue type 2 viruses associated with increased pathogenicity in the Americas," *Virology*, 230 (2), 244-251.

21. Rodriguez-Roche, R.M., Alvarez, M., Gritsun, T., Halstead, S., Kouri, G., Gould, E.A., *et al* (2005)," Virus evolution during a severe dengue epidemic in Cuba, 1997," *Virology*, 334 (2), 154-159.

22. Guzma'n, M.G., Kouri, G., Bravo, J., Valdes, L., Vazquez, S. and Halstead, S.B (2002), " Effect of age on outcome of secondary dengue 2 infections," *International Journal of Infectious Diseases*, 6 (2), 118-124.

23. Cologna, R. and Rico-Hesse, R (2003), "American genotype structures decrease dengue virus output from human monocytes and dendritic cells," *Journal of Virology*, 77 (7), 3929-3938.

24. Leitmeyer, K.C., Vaughn, D.W., Watts, D.M., Salas, R., Villalobos, I., Chacon, de, *et al* (1999), "Dengue virus structural differences that correlate with pathogenesis," *Journal of Virology*, 73 (6), 4738-4747.

25. Jessie, K., Fong, M.Y., Devi, S., Lam, S.K. and Wong. K.T (2004), "Localization of dengue virus in naturally infected human tissues, immunohistochemistry and in situ hybridization," *Journal of Infectious Diseases*, 189 (8), 1411-1418.

26. Paes, M.V., Pinhao, A.T., Barreto, D.F., Costa, S.M., Oliveira, M.P., Nogueira, A.C., *et al* (2005), " Liver injury and viremia in mice infected with dengue-2 virus," *Virology*, 338 (2), 236-246.

27. Seneviratne, S.L., Malavige, G.N. and de Silva, H.J (2006), " Pathogenesis of liver involvement during dengue viral infections," *Transactions of the Royal Society of Tropical Medicine and Hygiene*, 100 (7), 608-614.

28. Nkhoma, E.T., Poole, C., Vannappagari, V., Hall, S.A. and Beutler, E (2009), "The global prevalence of glucose-6-phosphate dehydrogenase deficiency : a systematic review and meta-analysis," *Blood Cells Molecules and Diseases*, 42 (3), 267-278.

29. Wu, Y.H., Tseng, C.P., Cheng, M.L., Ho, H.Y., Shih, S.R. and Chiu, D.T (2008), "Glucose-6phosphate dehydrogenase deficiency enhances human coronavirus 229E infection," *Journal ofInfectious Diseases*, 197 (6), 812-816.

30. Zivna, I., Green, S., Vaughn, D.W., Kalayanarooj, S., Stephens, H.A., Chandanayingyong, D., *et al* (2002), "T-cell responses to an HLA-B*07-restricted epitope on the dengue NS3 protein correlate With disease severity," *Journal of Immunology*, 168 (11), 5959-5965.

31. Polizel, J.R., Bueno, D., Visentainer, J.E., Sell, A.M., Borelli, S.D., Tsuneto, L.T., *et al* (2004), "Association of human leukocyte antigen DQ1 and dengue fever in a white Southern Brazilian

population," Memo'rias do Instituto Oswaldo Cruz, 99 (6), 559-562.

32. Alagarasu, K., Mulay, A.P., Sarikhani, M., Rashmika, D., Shah, P.S. and Celilia, D (2013), " Profile of human leukocyte antigen class I alleles in patients with dengue infection from Western India," *Human Immunology*, 74 (12), 1624-1628.

33. Malavige, G.N., Fernando, S., Fernando, D.J. and Seneviratne, S.L (2004), "Dengue viral infection, " *Postgraduate Medical Journal*, 80 (948), 588-601.

34. Rathakrishnan, A., Klekamp, B., Wang, S.M., Komarasamy, T.V., Natkunam, S.K., Sanchez-Anguiano, A, *et al* (2014), "Clinical and immunological markers of dengue progression in a study

cohort from a hyperendemic area in Malaysia," *PLoS ONE*, 9 (3), e92021. DOI: 10.1371/journal.pone.0092021

35. Cardozo, D.M., Moliterno, R.A., Sell, A.M., Guelsin, G.A.S., Beltrame, L.M., Clemintino, S.L., *et al* (2014), "Evidence of HLA-DQB1 contribution to susceptibility of dengue serotype 3 in dengue patients in southern Brazil," *Journal of Tropical Medicine*, Article ID 968262, 6 pages.

36. Alencar, L.X.E.de, Braga-Neto, U.deM., Nascimento, E.J.M.do, Cordeiro, M.T., Silva, A.M., Brito, A.A.de, *et al* (2013), "HLA-B*44 is associated with dengue severity caused by DENV-3 in a Brazilian population," *Journal of Tropical Medicine*, Article ID 648475, 11 pages.

37. Monteiro, S.P.,Brasil, P.E.A.A.do, Cabello, G.M.K., Souza, R.V.de, Brasil, P., Georg, I., *et al* (2012), "HLA-A*01 allele : a risk factor for dengue hemorrhagic fever in Brazil's population," *Memo'rias do Instituto Oswaldo Cruz, Rio de Janeiro*, 107 (2), 224-230.

38. Brown, M.G., Salas, R.A., Vikers, I.E., Heslop, O.D. and Smikle, M.F (2011), " Dengue HLA associations in Jamaicans," *West Indian Medical Journal*, 60 (2), 126-131.

39. Malavige, G.N., Rostron, T., Rohanachandra, L.T., Jayaratne, S.D., Fernando, N., Silva, A.D.De, *et al* (2011), "HLA class I and class II associations in dengue viral infections in a Sri Lankan population," *PLoS ONE*, 6 (6), e20581. DOI : 10.1371/journal.pone.0020581

40. Appanna, R., Ponnampalavanar, S., See, L.L.C. and Sekaran, S.D (2010), "Susceptible and protective HLA class 1 allele against dengue fever and dengue hemorrhagic fever patients in a Malaysian population," *PLoS ONE*, 5 (9), e13029. DOI: 10.1371/journal.pone.0013029

41. Falco'n-Lezama, J.A., Ramos, C., Zuñiga, J., Jua'rez-Palma, L., Rangel-Flores, H., Garcia-Trejo, A.R., *et al* (2009), "HLA class I and II polymorphisms in Mexican Mestizo patients with dengue fever," *Acta Tropica*, 112 (2), 193-197.

42. Vejbaesya, S., Luangtrakool, P., Luangtrakool, K, Kalayanarooj, S., Vaughn, D.W., Endy, T.P., *et al* (2009), "TNF and LTA gene, allele, and extended HLA haplotype associations with severe dengue virus infection in ethnic Thais," *Journal of Infectious*

Diseases, 199 (10), 1442-1448.

43. Lan, N.T.P., Kikuchi, M., Huong, V.T.Q., Ha, D.Q., Thuy, T.T., Tham, V.D., *et al* (2008), "Protective and enhancing HLA alleles, HLA-DRB1*0901 and HLA-A*24, for severe forms of dengue virus infection, dengue hemorrhagic fever and dengue shock syndrome," *PLoS Neglected Tropical Diseases*, 2 (10), e304. DOI: 10.1371/journal.pntd.0000304

44. Sierra, B., Alegre, R., Pe'rez, A.B., Garcia, G., Sturn-Ramirez, K., Obasanjo, O., *et al* (2007), "HLA-A, -B, -C, and -DRB1 allele frequencies in Cuban individuals with antecedents of dengue 2 disease : advantages of the Cuban population for HLA studies of dengue virus infection," *Human Immunology*, 68 (6), 531-540.

45. Likitnukul, S., Prappal, N., Pongpunlert, W., Kingwatanakul, P. and Poovorawan, Y (2004), " Dual infections : dengue hemorrhagic fever with unusual manifestations and mycoplasma pneumonia in a child," *Southeast Asian Journal of Tropical Medicine and Public Health*, 35 (2), 399-402.

46. Ali, F., Saleem, T., Khalid, U., Mehwood, S.F. and Jamil, B (2010), "Crimen-Congo hemorrhagic fever in a dengueendemic region: lessons for the future," *Journal of Infection in Developin Countries*, 4 (7), 459-463.

47. Wang, C.C., Wu, C.C., Liu, J.W., Lin, A.S., Liu, S.F., *et al* (2007), "Chest radiographic presentation in patients with dengue hemorrhagic fever," *American Journal of Tropical Medicine and Hygiene*, 77 (2), 291-296.

48. Mohamed, N.A., El-Raoof, E.A. and Ibraheem, H.A (2013), " Respiratory manifestations of dengue fever in Taiz-Yemen," *Egypt Journal of Chest Diseases and Tuberculosis*, 62 (NA), 319-323.

49. Asghar, J. and Farooq, K (2011), "Radiological appearance and their significance in the management of dengue hemorrhagic fever," *Pakistan Journal of Medical and Health Sciences*, 5 (4), 685-692.

50. Rodrigues, R.S., Brum, A.L.G., Paes, M.V., Po'voa, T.F., Basiliode-Oliveira, C.A., Marchiori, E., *et al* (2014), "Lung in dengue : computed tomography findings," *PLoS ONE*, 9 (5), e96313. DOI : 10.1371/journal.pone.0096313 51. Duchin, J.S., Koster, F.T., Peters, C.J., Simpson, G.L., Tempest, B., Zaki, S.R., *et al* (1994), "The Hantavirus Study Group. Hantavirus pulmonary syndrome : a clinical description of 17 patients with a newly recognized disease," *New England Journal of Medicine*, 330 (14), 949-955.

52. Castillo, C., Naranjo, J., Sepu'lveda, A., Ossa, G. and Levy, H (2001), "Hantavirus pulmonary syndrome due to Andes virus in Temuco, Chile : clinical experience with 16 adults," *Chest*, 120 (2), 548-554.

53. Vapalahti, O., Mustonen, J., Lundkvist. A., Henttonen, H., Plyusnin, A. and Vaheri, A (2003), "

Hantavirus infections in Europe," *Lancet Infectious Diseases*, 3 (10), 653-661.

54. Rasmuson, J., Pourazar, J., Linderholm, M., Sandström, T., Blomberg, A. and Ahlm, C (2011), " Presence of activated airway T lymphocytes in human Puumala hantavirus disease," *Chest*, 140 (3), 715-722.

55. Guirakhoo, F., Kitchener, S., Morrison, D., Forrat, R., McCarthy, K., Nicholas, R., *et al* (2006), "Live attenuated chimeric yellow fever dengue type 2 (ChimeriVax-DEN2) vaccine : Phase Iclinical trial for safety and immunogenicity : effect of yellow fever pre-immunity in induction of cross neutralizing antibody responses to all 4 dengue serotypes," *Human Vaccine*, 2 (2), 60-67.

56. Durbin, A.P., Whitehead, S.S., McArthur, J., Perreault, J.R., Blaney, J.E. Jr., Thumar, B., *et al*(2005), "rDEN4 delta 30, a live

attenuated dengue virus type 4 vaccine candidate, is safe, immunogenic, and highly infectious in healthy adult volunteers," *Journal of Infectious Diseases*, 191 (5), 710-718.

57. Raviprakash, K., Apt, D., Brinkman, A., Skinner, C., Yang, S., Dawes, G., *et al* (2006), "Achimeric tetravalent dengue DNA vaccine elicits neutralizing antibody to all four virus serotypes in rhesus macaques," *Virology*, 353 (1), 166-173.

58. Hermida, L., Bernardo, L., Martin, J., Alvarez, M., Prado, I., Lo', C., *et al* (2006), "Arecombinant fusion protein containing the domain III of the dengue-2 envelope protein is immunogenic and protective in nonhuman primates," *Vaccine*, 24 (16), 3165-3171.

59. Whitehead, S.S., Falqout, B., Hanley, K.A., Blaney, J.E. Jr., Markoff, L. and Murphy, BR (2003), " A live, attenuated dengue virus type 1 vaccine candidate with a 30-nucleotide deletion in the 3' untranslated region is highly attenuated and immunogenic in monkeys," *Journal of Virology*,77 (2), 1653-1657.

60. Edelman, R., Wasserman, S.S., Bodison, S.A., Putnak, R.J., Eckels, K.H., Tang, D., *et al* (2003), "Phase I trial of 16 formulations of a tetravalent live-attenuated dengue vaccine," *American Journal of Tropical Medicine and Hygiene*, 69 (6 Suppl), 48-60.