Abstract

The present study was conducted to evaluate drug resistance patterns of *Salmonella enterica* serotype *typhi* and *paratyphi* in enteric fever.

This study is an observational study.

The place of this study is the Department of Medicine, Liaquat University of Medical and Health Sciences Hospital and the duration from June 2012 to March 2013.

A sample of sixty seven patients was selected according to clinical and laboratory criteria. Venous blood samples of 5-10 ml were collected from antecubital veins and inoculated on blood culture bottles. Subcultures were also performed. Drug sensitivity patterns of salmonellae were tested according to standard method. Data was analyzed on SPSS version 21.0 using appropriate statistical testing.

Of sixty seven, 41(61.1%) were male and 26 (38.8%) female. The male to female ratio was 1.57:1. The mean age of subjects was 43±7.5 years. *Salmonella enterica* serotype *typhi* was isolated from 60 (89.5%), salmonellae *enterica* serotype *typhi* and salmonellae *enterica* serotype *paratyphi A* and *B* were found in 3 (4.4%) and 4 (5.9%) respectively. The Widal test yielded antibody *titers* of ≥1:160 and <1:160 in 57 (85%) and 10 (14.9%) respectively. The typhidot test was positive in 65 (97.5%) of patients. The multidrug resistance to first line drugs was observed in 53 (79.1%). Least drug resistance was found for *cefixime* 2 (2.7%) and highest for the chloramphenicol 56 (83.6%).

The present study observed drug resistant and multi-drug resistant *S. enterica* serotype *typhi* and *S. enterica* serotype *paratyphi A* and *B*.

Keywords: Drug sensitivity Enteric fever *Salmonellae typhi/paratyphi*.
Introduction

Enteric fever is commonly known as typhoid fever. Salmonella enterica serotype typhi and salmonellae enterica serotype paratyphi are most common etiological agents of enteric fever. The term typhoid fever applies when serotype *typhi* is causative agent. Salmonellae are transmitted by contaminated food, milk, water or drinks. Clinical manifestations vary from person to person. The incubation period varies from 5 to 14 days (Schwartz et al 2012) (Ochiai et al 2008). Pakistan is hyper-endemic area for enteric fever. Its incidence was 412 per million in children of 5-15 years (Ochiai et al 2008). The isolation of *Salmonellae enterica* serotype *typhi/paratyphi* from blood, bone marrow and urine is necessary for definitive diagnosis; however bone marrow cultures are regarded as 'gold standard method'. The Widal and typhidot tests are alternative serology methods for diagnosis (Behrmann et al 2008). The drug resistance of salmonellae was rare prior to 1980s. The treatment of choice was chloramphenicol, but emergence of resistance in mid 1980s, created therapeutic problems particularly in countries where enteric fever is endemic. Then the ciprofloxacin became drug of choice for typhoid (Somily 2010). The *S. enterica* serotype *typhi* resistant to chloramphenicol, ampicillin, and co-trimoxazole were reported from South America, Africa, India and Saudi Arabia (Somily 2010). Simultaneously, Southern Vietnam reported multi-drug resistance isolates in early 1990s (Somily 2010). Shortly after, isolates of *S. enterica* serotype *typhi* resistant to quinolone were reported from same geographical areas with resistance to first line drugs. This was a major threat for treatment of typhoid fever. Isolates of *S. enterica* serotype *typhi* resistant to nalidixic acid and ciprofloxacin increased to 23% in United Kingdom in 1999s. Somily (2010) reported drug resistant isolates of *S. enterica* serotype *typhi* in patient’s returning from travel to Asia (Somily 2010). The quinolones and 3rd generation cephalosporins are reported of having successful cure rates of 82-97% and 100% respectively (Hazir 2010). But a number of resistance mechanisms to quinolone in certain parts of the world led to decreased susceptibility of *S. enterica* serotype *typhi* to these drugs also (Somily 2010). Like many tropical areas, enteric fever is hyper-endemic in Pakistan. It is major public health problem and a cause of mortality in Sindh. The present study was aimed to have insight into current trends of drug sensitivity and resistance patterns of *S. enterica* serotype *typhi* and salmonellae *enterica* serotype *paratyphi* in Sindh province.

Methodology

Sixty seven adult patients with a clinical diagnosis suggestive of enteric fever, admitted at Department of Medicine, Liaquat University of Medical and Health Sciences Hospital from June 2012 to March 2013, were enrolled in the study. The clinical history of symptoms and drugs was sought from the patients. Enteric fever was defined according to gold standard method of isolation of *S. enterica* serotype *typhi* and salmonellae *enterica* serotype *paratyphi* A and B on blood culture. (Konemann 1997). Venous blood samples were collected from febrile patient whether taking or not an antibiotic therapy. Complete blood counts, Widal test, typhoid test and blood cultures were carried out according to standard methods by hematologist. Bottles containing 50 ml of brain heart infusion broth were inoculated with blood for culture. The samples were processed according to standard recommended techniques for blood culturing. (Konemann 1997). The blood cultures were incubated at 37°C for seven days and bacterial growth was observed on daily basis. Subcultures were performed on 1st, 3rd, 5th and 7th day of incubation on 5% sheep blood agar and Mac Conkey medium (Kumar 2008) Non-lactose fermenting colonies were screened by biochemical reactions, and further identified by serology. Antibiotic sensitivity and resistance of microorganisms were tested according to standardized Kirby–Bauer disc diffusion method. (Bauer 1966). Antimicrobial susceptibility patterns were determined using commercial antimicrobial disks (Oxoid, Basingstoke, UK). Antimicrobial susceptibility testing
against chloramphenicol (30ug), co-
trimoxazole (25ug), ampicillin (10ug),
amoxicillin (10ug), amoxi-clav (20/10ug),
ciprofloxacin (5 ug), cefotaxime (10ug),
ceftriaxone (10ug), levofloxacin (5ug), and
cefixime (10ug) performed by Clinical
Laboratory Standard Institute (CLSI 2010).
The study was approved by the ethics
committee of institute. Data was analyzed
on SPSS version 21.0. using the chi-square
and student’s t-tests for categorical and
continuous variables respectively.

Results

The mean age of subjects was 43±7.5 years.
Of sixty seven adult subjects, 41(61.1%)
were male and 26 (38.8%) were female.
(p=0.001) The male to female ratio was
1.57:1. The demographic characteristics of
patients and clinical symptoms and signs
are summarized in table I.

Clinical Features: The most frequent
clinical symptoms and signs were fever,
chills headache and anorexia. The bloody
diarrhea was also observed. The
hepatosplenomegaly were noted. The

Antimicrobial drugs taken as self
medication was noted in twenty nine
(43.2%) of patients; and it was also noted
frequency and % of clinical symptoms and
signs are shown in table I. Out 67 subjects,
leukocytosis and leukopenia were noted in
31 (46.2%) and 29 (43%) respectively.

Isolation Rate: The S.enterica serotype
typhi, S. enterica serotype typhi/paratyphi
and S.enterica serotype paratyphi A and B
was isolated from 60 (89.5%), 3(4.4%) and 4
(5.9%) respectively (p=0.001).

Serological Tests: Typhoid test was
positive in 65 (97.5%) of patients. Antibody O
titers of ≥1:160 and <1:160 were detected in
57 (85%) and 10 (14.9%) of patients respectively.

Drug Resistance Patterns: The
antibiotic sensitivity and resistance
patterns are shown in table II. The
multidrug resistance to first line drugs
was observed in 53 (79.1%). The highest
and lowest drug sensitivity were noted
for cefixime and chloramphenicol in 65
(97.3%) and 11 (16.4%) of patients
respectively. Least drug resistance was
found for cefixime 2 (2.7%) and highest
for chloramphenicol 56 (83.6%).
that the patients were running fever and
were found blood cultures positive in
59.7%.

Table I. Demographic Data and Clinical Features of Study Population (N=67)

<table>
<thead>
<tr>
<th></th>
<th>No. of cases</th>
<th>%</th>
</tr>
</thead>
<tbody>
<tr>
<td>Male (M)</td>
<td>41</td>
<td>61.1</td>
</tr>
<tr>
<td>Female (F)</td>
<td>26</td>
<td>38.8</td>
</tr>
<tr>
<td>Fever</td>
<td>67</td>
<td>100</td>
</tr>
<tr>
<td>Chills</td>
<td>49</td>
<td>73.1</td>
</tr>
<tr>
<td>Headache</td>
<td>53</td>
<td>79.1</td>
</tr>
<tr>
<td>Sore throat</td>
<td>27</td>
<td>40.2</td>
</tr>
<tr>
<td>Coated tongue</td>
<td>32</td>
<td>47.7</td>
</tr>
<tr>
<td>Anorexia</td>
<td>65</td>
<td>97</td>
</tr>
<tr>
<td>Vomiting</td>
<td>21</td>
<td>31.3</td>
</tr>
<tr>
<td>Bloody diarrhoea</td>
<td>11</td>
<td>16.4</td>
</tr>
<tr>
<td>Rose spots</td>
<td>13</td>
<td>19.4</td>
</tr>
<tr>
<td>Meningism</td>
<td>21</td>
<td>31.3</td>
</tr>
<tr>
<td>Hepatomegaly</td>
<td>05</td>
<td>07.4</td>
</tr>
<tr>
<td>Splenomegaly</td>
<td>19</td>
<td>28.3</td>
</tr>
<tr>
<td>Respiratory symptoms</td>
<td>31</td>
<td>46.2</td>
</tr>
</tbody>
</table>
Table II. Antibiotic Sensitivity and Resistance Patterns of Isolated Salmonellae

<table>
<thead>
<tr>
<th>Drug</th>
<th>Sensitive (%)</th>
<th>Resistance (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Chloramphenicol (30 µg)</td>
<td>11 (16.4%)</td>
<td>56 (83.5%)</td>
</tr>
<tr>
<td>Co-trimoxazole (25 µg)</td>
<td>13 (19.4%)</td>
<td>54 (80.5%)</td>
</tr>
<tr>
<td>Ampicillin (10 µg)</td>
<td>12 (17.9%)</td>
<td>55 (82.0%)</td>
</tr>
<tr>
<td>Amoxicillin (10 µg)</td>
<td>17 (25.3%)</td>
<td>50 (74.6%)</td>
</tr>
<tr>
<td>Amoxi-Clav (20/10 µg)</td>
<td>26 (38.8%)</td>
<td>41 (61.1%)</td>
</tr>
<tr>
<td>Ciprofloxacin (5 µg)</td>
<td>23 (34.3%)</td>
<td>44 (65.6%)</td>
</tr>
<tr>
<td>Cefotaxime (10 µg)</td>
<td>33 (49.2%)</td>
<td>34 (50.7%)</td>
</tr>
<tr>
<td>Ceftriaxone (10 µg)</td>
<td>51 (76.1%)</td>
<td>16 (23.8%)</td>
</tr>
<tr>
<td>Levofloxacin (5 µg)</td>
<td>61 (91.0%)</td>
<td>6 (8.9%)</td>
</tr>
<tr>
<td>Cefixime (10 µg)</td>
<td>65 (97.3%)</td>
<td>2 (2.9%)</td>
</tr>
</tbody>
</table>

Discussion

The multi-drug resistance defined as resistance to first line drugs i.e. chloramphenicol, cotrimoxazole, and ampicillin, was noted in 79.1% of *S. enterica* serotype *typhi/paratyphi* in present study. The results are comparable to previous studies which have reported 80-90% (Hatta 2008) and 100% (Misra 2005). The increased drug resistance results from the exploitation of drug use by chemist, druggist, quacks, and paramedics and more over by patients himself as self medication. The access to drugs is very easy in our country because drug prescription and sale is uncontrolled by law due to prevalent political, social and other issues (Siddiqui et al 2006) (Abdullah 2012). The regulations of drug prescription and sale are not obliged by pharmacists. This results in injudicious administration and sub-optimal drug dosaging which have contributed much to the reduced susceptibility and emergence of multi drug resistant *S. enterica* serotype *typhi/paratyphi*, not only to the first line drugs but also to the cephalosporins, quinolones, and macrolides. (Siddiqui et al 2006) (Abdullah 2012).

One study from Indonesia reported that up to 90% of enteric fever patients were receiving antimicrobials as self medication at home and bed rest; (Hatta 2008) other patients, those with more severe disease and those treated at home who then developed persisting complaints, presented at the health care centers and hospitals (Tjiandi 2003). The similar conditions of drug usage are prevailing in our society. A recent study has reported overall multi drug resistance in Salmonellae in enteric fever of 62.64% (Abdullah 2012) which is low but yet comparable to our present study. The recently reported amoxicillin resistance of 96.4% is higher as we found 74.6%, and co-trimoxazole resistance of 29.91% is very low as compared to 80.5% in our present study (Abdullah 2012). We found cefixime, levofloxacin and ceftriaxone sensitivity of 97.3%, 91% and 76.1% respectively. Our findings are comparable to previous studies (Siddiqui et al 2006) (Kumar et al 2008) (Abdullah 2012). Our multi-drug resistance finding of 79.1% is comparable to previous reports from Nigeria of 80% (Akineymi et al 2007) and Pakistan (Hassan et al 2008) (Butt et al 2005). Possibly due to socio-economic constraints and indiscriminate drug use, the prevalence of quinolone resistance has gradually increased (Mubeena et al. 2000) (Fangtham et al 2008). The first case of fluoroquinolones treatment failure was reported in Pakistan in 1993 (Butt et al 2005). We found drug resistance salmonellae to ciprofloxacin and levofloxacin of 65.6% and 8.9% respectively. A study from Agha Khan University reported multidrug resistant *S. enterica* serotype *typhi* in 66% of children from Karachi (Khan et al 2012). Similarly a study from India reported that out of 107 salmonellae isolates, 94 (87.8%) were nalidixic acid-resistant (NAR) *Salmonella* and 36 were resistant to azithromycin. Among the 57 treatment
histories, 19 (33%) patients had documented clinical non-response to azithromycin which required change of therapy (Rai et al 2012). This is seemingly an alarming situation showing development of drug resistant salmonellae against macrolides, quinolone and fluoroquinolones which may increase the resistance in near future. Fluoroquinolones are under strict prescription of clinicians in advanced countries (Fangtham et al 2008) but freely available in developing countries like Pakistan (Abdullah 2012). In our study the crucial finding is that the twenty nine (43.2%) of patients taking antibiotic therapy yielded positive blood cultures in 59.7% of them, this reveals the existence of drug resistant salmonellae in the community. The easy access of public to the prescription drugs is an illegal practice and must be controlled and this may help to prevent widespread drug resistant salmonellae in future.

Conclusion

The present study observed drug resistant and multi-drug resistant S. enterica serotype typhi/paratyphi A& B in majority of enteric fever patients. The increasing drug resistant salmonellae must be controlled to prevent epidemics of enteric fever within community.

References


